## **Research Brief**



## Evaluation of an alcohol-based antiseptic for nasal decolonization of methicillin-resistant *Staphylococcus aureus* in colonized patients

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Mupirocin is commonly used for nasal decolonization of *Staphylococcus aureus*, but the emergence of resistance is a growing concern.<sup>1–3</sup> Alcohol-based nasal antiseptics are a potential alternative to mupirocin. Steed et al<sup>4</sup> reported that an alcohol-based nasal antiseptic reduced the burden of *S. aureus* in the nares of health-care personnel. Moreover, in a quasi-experimental study, perioperative use of an alcohol-based nasal antiseptic by orthopedic patients and personnel was associated with reduced infection rates.<sup>5</sup> These findings are promising, but more information is needed regarding the efficacy of alcohol-based sanitizers in reducing nasal *S. aureus* in colonized patients.

We conducted 2 nonblinded randomized trials to determine the effectiveness of a single application of 2 different doses of a commercial alcohol-based (ie, 62% ethanol) nasal sanitizer (Nozin Nasal Sanitizer antiseptic, Global Life Technologies, Chevy Chase, MD) to reduce methicillin-resistant *S. aureus* (MRSA) in colonized patients. Patients receiving systemic antibiotics or nasal antiseptics were excluded. In the first trial (N = 30 participants), a single saturated swab was used to apply the product or phosphate-buffered saline to both nasal vestibules. In the second trial (N = 40), 3 separate saturated swabs were used to apply consecutive treatments over 3 minutes. The manufacturer recommends the triple-dose application for preoperative treatment, and twice-daily application of a single saturated swab is recommended postoperatively and for intensive care unit decolonization. One individual (A.K.) performed the nasal applications according to the manufacturer's instructions.

For all participants, rayon swabs (BBL Culture Swabs, Becton Dickinson, Franklin Lakes, NJ) were used to collect cultures from the anterior nares and vestibule prior to and 10 minutes, 2 hours, and 6 hours after application. Swabs were also used to collect cultures for MRSA from the clothing and skin (hands, groin, and chest/axilla) for the initial 23 participants. The swabs were cultured for MRSA as previously described.<sup>6</sup> Lab personnel were blinded to the study groups.

We also applied 3 single-dose applications of the sanitizer to the nares of 9 MRSA-colonized patients over 8 hours (applications at 0, 4, and 8 hours) using the protocol of Steed et al.<sup>4</sup> Nares cultures were collected at baseline and 2 hours after the third dose.

Repeated measures analysis of variance was used to compare treatment and control groups. For the repeated dosing group, MRSA concentrations during treatment were compared to pretreatment concentrations. Analyses were performed using R version 3.5.1 statistical software (R Foundation for Statistical Computing, Vienna, Austria). The study protocol was approved by the facility's institutional review board.

For the single-application trials, 68 of 70 (97%) participants were male, and the mean age was 65. The single-dose application was associated with a nonsignificant trend toward reduced mean MRSA concentrations in the treatment group versus the control group at 10 minutes and at 2 hours after dosing (Fig. 1A). The triple-dose application significantly reduced mean MRSA concentrations in comparison to controls at 10 minutes and at 2 hours after dosing (Fig. 1B). MRSA was frequently recovered from the clothing (19 of 23, 83%) and skin (17 of 23, 74%) of participants.

For the assessment of repeated dosing of the alcohol-based nasal sanitizer over 8 hours, all 9 participants were male. There was no reduction in MRSA 2 hours after the final dose in comparison to baseline (mean  $\pm$ SE, 2.3  $\pm$ 0.43 vs 2.3  $\pm$ 0.78 log<sub>10</sub> colony-forming units per swab; *P* > .05).

In summary, single-dose applications of an alcohol-based sanitizer did not significantly reduce nasal MRSA, and a triple-dose application only transiently reduced the burden of MRSA with no significant reduction by 6 hours after dosing. Several factors might contribute to the relatively limited efficacy of the alcoholbased nasal sanitizer in MRSA-colonized patients. First, adequate application of alcohol in the anterior nares is relatively challenging. Second, organic material reduces the efficacy of alcohol and may be present in relatively large amounts in the nares.<sup>7,8</sup> Third, *S. aureus* in the nares may not be accessible to alcohol treatment (ie, embedded within mucus and at the base of hair follicles).<sup>9</sup> At such sites, *S. aureus* in the nares may be analogous to resident skin flora on hands that is minimally suppressed by alcohol.<sup>7</sup> Finally, alcohol has only a transient antibacterial effect, and more persistent activity may be required to reduce the burden of nasal MRSA.<sup>7</sup>

Our results differ from a recent report of significant reductions in the burden of nasal *S. aureus* in healthcare personnel with the alcohol-based nasal antiseptic applied 3 times during an 8-hour period.<sup>4</sup> The differing results may be related in part to differences between colonized patients and personnel. Most of the patients we studied were persistent carriers, whereas some healthcare

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**Fig. 1.** Effect of a single application of an alcohol-based nasal sanitizer versus phosphate-buffered saline on the burden of methicillin-resistant *Staphylococcus aureus* (MRSA) in the nares of colonized patients. (A) Single application (N = 30 participants). (B) Triple application with 3 consecutive treatments over 3 minutes (N = 40 participants). \*P < .05. Note. CFU, colony-forming units.

personnel in the study of Steed et al<sup>4</sup> may have been transient carriers. Many of the colonized patients also had a high burden of contamination on their clothing and skin, providing a potential reservoir for recurrent inoculation of the nares.<sup>10</sup>

Our study has some limitations. The study was nonblinded regarding the application of the control or alcohol solution, and it was conducted in a single center with primarily male participants. For the assessment of repeated applications, no control group was included, and the number of participants was low. Because the triple-dose alcohol application significantly reduced MRSA for at least 2 hours, it is possible that frequent, relatively high doses of alcohol might maintain *S. aureus* suppression. If contamination on clothing and skin contributes substantially to recolonization of the nares, it is possible that alcohol-based nasal sanitizers might be more effective in combination with chlorhexidine bathing.

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## References

1. McDanel JS, Murphy CR, Diekema DJ, et al. Chlorhexidine and mupirocin susceptibilities of methicillin-resistant Staphylococcus aureus from

colonized nursing home residents. Antimicrob Agents Chemother 2013; 57:552-558.

- Huang SS, Septimus E, Kleinman K, et al. Chlorhexidine versus routine bathing to prevent multidrug-resistant organisms and all-cause bloodstream infections in general medical and surgical units (ABATE Infection trial): a cluster-randomised trial. *Lancet* 2019;393:1205–1215.
- Huang SS, Singh R, McKinnell JA, et al. Decolonization to reduce postdischarge infection risk among MRSA carriers. N Engl J Med 2019;380: 638–650.
- Steed LL, Costello J, Lohia S, Jones T, Spannhake EW, Nguyen S. Reduction of nasal Staphylococcus aureus carriage in health care professionals by treatment with a nonantibiotic, alcohol-based nasal antiseptic. *Am J Infect Control* 2014;42:841–846.
- Mullen A, Wieland HJ, Wieser ES, Spannhake EW, Marinos RS. Perioperative participation of orthopedic patients and surgical staff in a nasal decolonization intervention to reduce Staphylococcus spp. surgical site infections. *Am J Infect Control* 2017;45:554–556.
- Sunkesula V, Kundrapu S, Macinga DR, Donskey CJ. Efficacy of alcohol gel for removal of methicillin-resistant Staphylococcus aureus from hands of colonized patients. *Infect Control Hosp Epidemiol* 2015;36:229–231.
- Boyce, JM, Pittet, D. Guideline for hand hygiene in health-care settings: recommendations of the Healthcare Infection Control Practices Advisory Committee and the HICPAC/SHEA/APIC/IDSA Hand Hygiene Task Force. *Infect Control Hosp Epidemiol* 2002;23:S3–S40.
- Loveday, HP, Wilson, JA, Pratt, RJ, et al. Epic3: national evidence-based guidelines for preventing healthcare-associated infections in NHS hospitals in England. J Hosp Infect 2014;86:S1–S70.
- Ten Broeke-Smits NJ, Kummer JA, Bleys RL, Fluit AC, Boel CH. Hair as a niche of Staphylococcus aureus in the nose: is a more effective decolonization strategy needed? J Hosp Infect 2010;76:211–214.
- Kanwar A, Cadnum JL, Thakur M, Jencson AL, Donskey CJ. Contaminated clothing of methicillin-resistant Staphylococcus aureus (MRSA) carriers is a potential source of transmission. Am J Infect Control 2018;46:1414–1416.